



Estimation of Genetic Parameters for Yield Traits in Greengram (*Vigna radiata* (L.) Wilczek) Mutants under EMS Mutagenesis of M3 Generation

V. Rohini¹, K. Srinivas Naik², Deepika Dasyam² and Sudhakar Poda^{3*}

¹Assistant Professor of Biotechnology, Department of Biotechnology, Government Degree College Siddipet, Telangana, India.

²Centre for Plant Molecular Biology, Osmania University, Hyderabad, Telangana, India.

³Department of Biotechnology, Acharya Nagarjuna University, Nagarjuna Nagar, Guntur, Andhra Pradesh, India.

*Corresponding author email id: sudhakarpodha@gmail.com

Date of publication (dd/mm/yyyy): 10/04/2026

Abstract – The current study was carried out with mutants of greengram (WGG-42) genotype collected from from ICAR-CRIDA, Santoshnagar, Hyderabad and the work was done at the Center for Plant Molecular Biology at Osmania University in Hyderabad and the Department of Genetics and Plant Breeding at Acharya N.G. Ranga Agricultural University in Lam, Guntur, Andhra Pradesh. The aim of the study is to analyse genetic parameter for yield characters in M3 generation. The results showed the correlation analysis revealed that number of secondary branches per plant, number of clusters per plant, number of pods per plant, number of seeds per pod, and 100-seed weight exhibited strong positive associations with seed yield, indicating their direct contribution to productivity. The results also showed the estimates of heritability and genetic advance showed that plant height, number of primary and secondary branches per plant, number of pods per plant, number of seeds per plant, and seed yield per plant recorded high heritability coupled with high genetic advance, suggesting the predominance of additive gene action. These traits offer substantial scope for improvement through direct selection in mutant populations. In contrast, traits such as 100-seed weight and days to maturity exhibited moderate heritability with low to moderate genetic advance, implying the influence of non-additive gene effects and environmental factors, thereby requiring recurrent selection or hybridization strategies for effective improvement. The study confirmed that EMS-induced mutagenesis is an effective approach for generating genetic variability in green gram. The M₃ generation mutants displayed significant variability with promising heritable traits for yield enhancement.

Keywords – Green Gram, Correlation, Genetic Advance and Heritability.

I. INTRODUCTION

Greengram, also known as mungbean (*Vigna radiata* (L.) Wilczek), is one of the most important pulse crops cultivated across tropical and subtropical regions of Asia, Africa, and parts of South America. It belongs to the family Fabaceae and is valued for its high nutritional content, short life cycle, and ability to enrich soil fertility through biological nitrogen fixation. The seeds contain approximately 24-26% protein, 1-1.5% fat, 60–65% carbohydrates, and are rich in essential amino acids such as lysine and leucine, making them a key dietary component in vegetarian populations (Kaur et al., 2023). A key objective of mutation breeding is to enhance genetic variability-the raw material for selection and crop improvement. Quantitative genetic parameters such as genotypic and phenotypic coefficients of variation (GCV, PCV), heritability, and genetic advance provide insights into the extent of variability and the potential for selection in mutant populations (Falconer & Mackay, 2023). High heritability coupled with high genetic advance indicates additive gene action and suggests that selection would be effective for the trait (Basavaraj et al., 2022). Moreover, correlation studies among traits help



in understanding the interrelationships between yield and its contributing components, guiding breeders to select indirectly for yield improvement through easily measurable traits (Chaudhary et al., 2024).

II. MATERIAL AND METHODS

The present investigation was carried out at the Department of Genetics and Plant Breeding, Acharya N.G. Ranga Agricultural University, Lam, Guntur, Andhra Pradesh and Centre for Plant Molecular Biology, Osmania University, Hyderabad during Sept, 2025 to Dec, 2025. The greengram (*Vigna radiata* (L.) Wilczek) genotype-WGG-42, obtained from ICAR-CRIDA, Santoshnagar, Hyderabad and the ethyl methane sulphonate (EMS) was obtained from HIMEDIA Laboratory limited, Mumbai, India, was used as chemical mutagen to induce the blackgram seeds. The aim of the study is to analyses genetic parameter for yield characters in M3 generation.

Phenotypic and genotypic correlations were worked out using the formulae suggested by Johnson et al. (1955). Significance of correlation coefficients was tested by comparing phenotypic and genotypic correlation coefficients with the correlation table values given by Fisher and Yates (1963) at (n-2) degrees of freedom where 'n' denotes the number of paired observations used in the calculation. Phenotypic and genotypic coefficients of variation were calculated based on the method advocated by Burton (1952). Categorisation of the range of variation was followed as proposed by Sivasubramanian and Menon (1973). Heritability in broad sense was estimated as per Allard (1960) and expressed in percentage. Genetic advance was estimated by the method suggested by Johnson et al. (1955). Genetic advance was expressed as percentage of mean by using the following formula as given by Johnson et al. (1955).

III. RESULTS AND DISCUSSIONS

3.1. Correlation Analysis

The phenotypic correlation coefficients among yield and yield-related traits in M₃ generation of greengram mutants are presented in Table 1.

Table 1. Phenotypic correlation for seed yield and yield contributing traits in greengram mutants in M3 generations.

| | Pl. Ht (cm) | No. of P. Br./Pl | No. of S. Br./Pl | Days to F. Ini. | Days to P.M | No. L/Pl | No. Cl/Pl | No. P/Cl | No. P/Pl | No. S/PD | No S/Pl | 100 S. Wt (g) |
|-----------------|----------------|---------------------|---------------------|--------------------|----------------|-------------|--------------|-------------|-------------|-------------|------------|------------------|
| No. of P.Br./Pl | 0.963** | | | | | | | | | | | |
| No. of S.Br./Pl | 0.881** | 0.908** | | | | | | | | | | |
| Days to F. Ini. | -0.891** | -0.936** | -0.872** | | | | | | | | | |
| Days to P.M. | -0.892** | -0.893** | -0.881** | 0.873** | | | | | | | | |
| No. L/Pl | 0.970** | 0.979** | 0.864** | -0.933** | -0.893** | | | | | | | |
| No. Cl/Pl | 0.914** | 0.942** | 0.881** | -0.912** | -0.832** | 0.927** | | | | | | |
| No. P/Cl | 0.855** | 0.881** | 0.860** | -0.824** | -0.824** | 0.847** | 0.861** | | | | | |
| No. P/Pl | 0.906** | 0.932** | 0.903** | -0.898** | -0.848** | 0.903** | 0.980** | 0.926** | | | | |
| No. S/PD | 0.847** | 0.812** | 0.752** | -0.715** | -0.812** | 0.782** | 0.813** | 0.755** | 0.821** | | | |
| No S/Pl | 0.895** | 0.912** | 0.894** | -0.867** | -0.840** | 0.874** | 0.967** | 0.906** | 0.991** | 0.872** | | |
| 100 S. Wt(g) | 0.731** | 0.747** | 0.811** | -0.753** | -0.679** | 0.670** | 0.780** | 0.767** | 0.823** | 0.629** | 0.815** | |



| | Pl. Ht (cm) | No. of P. Br./Pl | No. of S. Br./Pl | Days to F. Ini. | Days to P.M | No. L/Pl | No. Cl/Pl | No. P/Cl | No. P/Pl | No. S/PD | No S/Pl | 100 S. Wt (g) |
|-------------|-------------|------------------|------------------|-----------------|-------------|----------|-----------|----------|----------|----------|---------|---------------|
| S.Y. (g)/Pl | 0.882** | 0.897** | 0.899** | -0.860** | -0.834** | 0.854** | 0.959** | 0.896** | 0.987** | 0.857** | 0.997** | 0.846** |

*Significant at 0.05% and ** 0.01 % level.

Pl. Ht (cm)-Plant Height (cm); No. of P.Br./Pl- Number of Primary Branches per Plant; No. of S.Br./Pl- Number of Secondary Branches per Plant; Days to F. Ini. - Days to Flower Initiation; Days to P.M - Days to Pod Maturity; No. L/Pl- Number of Leaves per plant; No. Cl/Pl- Number of Clusters per plant; No. P/Cl- Number of Pods per Cluster; No. P/Pl- Number of Pods per Plant; No. S/PD-Number of Seeds per Pod; No S/Pl- Number of Seeds per Plant; S.Y(g)/Pl- Seed Yield (g/pl); 100 S. Wt(g)-100 Seed Weight (g).

Seed yield per plant exhibited highly significant and positive phenotypic correlations with plant height ($r = 0.882^{**}$), number of primary branches per plant ($r = 0.897^{**}$), number of secondary branches per plant ($r = 0.899^{**}$), number of leaves per plant ($r = 0.854^{**}$), number of clusters per plant ($r = 0.959^{**}$), number of pods per cluster ($r = 0.896^{**}$), number of pods per plant ($r = 0.987^{**}$), number of seeds per pod ($r = 0.857^{**}$), number of seeds per plant ($r = 0.997^{**}$), and 100-seed weight ($r = 0.846^{**}$). It was negatively phenotypic correlated with days to flowering ($r = -0.860^{**}$) and days to maturity ($r = -0.834^{**}$).

Correlation studies provide insight into the degree and direction of association among different yield and yield-contributing traits, which helps in formulating effective selection strategies for yield improvement. Both phenotypic and genotypic correlations were estimated among various traits in Greengram (*Vigna radiata* L.) mutants of the M₃ generation.

Table 2. Phenotypic and Genotypic correlation for seed yield in greengram mutants in M3 generations.

| Characters | Seed Yield (g/pl) | |
|-----------------|------------------------|-----------------------|
| | Phenotypic Correlation | Genotypic Correlation |
| Pl. Ht(cm) | 0.882** | 0.912** |
| No. of P.Br./Pl | 0.897** | 0.930** |
| No. of S.Br./Pl | 0.899** | 1.000** |
| Days to F.Ini. | -0.860** | -0.912** |
| Days to P.M | -0.834** | -0.930** |
| No. L/Pl | 0.854** | 0.910** |
| No. Cl/Pl | 0.959** | 0.994** |
| No. P/Cl | 0.896** | 0.967** |
| No. P/Pl | 0.987** | 0.996** |
| No. S/PD | 0.857** | 0.947** |
| No S/Pl | 0.997** | 0.999** |
| 100 S. Wt(g) | 0.846** | 1.006** |

*Significant at 0.05% and ** 0.01 % level.

3.1.1. Phenotypic Correlation

The phenotypic correlation coefficients among different characters (Table ...) revealed that seed yield per pl-



-ant showed a highly significant and positive association with most of the yield-contributing traits such as number of primary branches per plant ($r = 0.930^{**}$), number of secondary branches per plant ($r = 1.000^{**}$), number of leaves per plant ($r = 0.910^{**}$), number of clusters per plant ($r = 0.994^{**}$), number of pods per cluster ($r = 0.967^{**}$), number of pods per plant ($r = 0.996^{**}$), number of seeds per pod ($r = 0.947^{**}$), number of seeds per plant ($r = 0.999^{**}$), and 100-seed weight ($r = 1.006^{**}$). This indicates that these traits are major determinants of yield and may be simultaneously improved through selection.

On the contrary, days to flowering initiation ($r = -0.912^{**}$) and days to physiological maturity ($r = -0.930^{**}$) exhibited a significant negative correlation with seed yield per plant. This suggests that early flowering and early maturity are advantageous traits for yield improvement, especially under short-duration cropping systems or stress environments.

The strong and significant positive correlations among number of clusters per plant, number of pods per plant, and seed yield imply that these traits are interdependent and contribute synergistically to productivity. Similar findings were reported by Gadakh et al. (2013), Konda et al. (2019), and Dudhe et al. (2021), who observed a positive association of branching and pod-bearing traits with seed yield in *Vigna radiata*.

3.1.2. Genotypic correlation

The genotypic correlation coefficients among yield and yield-related traits in M_3 generation of greengram mutants are presented in Table 2.

Table 3. Genotypic correlation for seed yield and yield contributing traits in greengram mutants in M_3 generation.

| | Pl. Ht(cm) | No. of P. Br./Pl | No. of S.Br./Pl | Days to F. Ini. | Days to P.M. | No. L/Pl | No. Cl/Pl | No. P/Cl | No. P/Pl | No. S/PD | No S/Pl | 100 S. Wt (g) |
|------------------|------------|------------------|-----------------|-----------------|--------------|----------|-----------|----------|----------|----------|---------|---------------|
| No. of P. Br./Pl | 0.979** | | | | | | | | | | | |
| No. of S. Br./Pl | 0.952** | 0.952** | | | | | | | | | | |
| Days to F. Ini. | -0.903** | -0.958** | -0.946** | | | | | | | | | |
| Days to P.M. | -0.972** | -0.995** | -0.988** | 0.965** | | | | | | | | |
| No. L/Pl | 0.988** | 0.999** | 0.947** | -0.944** | -0.993** | | | | | | | |
| No. Cl/Pl | 0.953** | 0.979** | 1.006** | -0.952** | -0.981** | 0.957** | | | | | | |
| No. P/Cl | 0.916** | 0.962** | 0.954** | -0.898** | -0.949** | 0.949** | 0.991** | | | | | |
| No. P/Pl | 0.923** | 0.953** | 0.994** | -0.927** | -0.948** | 0.932** | 0.998** | 0.989** | | | | |
| No. S/PD | 1.028** | 1.000** | 1.019** | -0.905** | -1.002** | 1.010** | 0.989** | 0.968** | 0.964** | | | |
| No S/Pl | 0.925** | 0.944** | 1.001** | -0.915** | -0.941** | 0.925** | 0.998** | 0.982** | 1.000** | 0.956** | | |
| 100 S. Wt(g) | 0.897** | 0.929** | 1.007** | -0.984** | -0.947** | 0.911** | 1.015** | 0.894** | 0.994** | 0.941** | 1.002** | |
| S.Y. (g)/Pl | 0.912** | 0.930** | 1.000** | -0.912** | -0.930** | 0.910** | 0.994** | 0.967** | 0.996** | 0.947** | 0.999** | 1.006** |

*Significant at 0.05% and ** 0.01 % level.

Pl. Ht(cm)- Plant Height (cm); v No. of P.Br./Pl- Number of Primary Branches per Plant; No. of S.Br./Pl- Number of Secondary Branches per Plant; Days to F.Ini.- Days to Flower Initiation; Days to P.M- Days to Pod Maturity; No. L/Pl- Number of Leaves per plant; No. Cl/Pl- Number of Clusters per plant; No. P/Cl- Number of Pods per Cluster; No. P/Pl- Number of Pods per Plant; No. S/PD- Number of Seeds per Pod; No S/Pl- Number of Seeds per Plant; S.Y(g)/Pl- Seed Yield (g/pl); 100 S. Wt(g)- 100 Seed Weight (g).



Seed yield per plant showed highly significant and positive genotypic correlations with almost all yield-related traits such as plant height ($r = 0.912^{**}$), number of primary branches ($r = 0.930^{**}$), number of secondary branches ($r = 1.000^{**}$), number of leaves ($r = 0.910^{**}$), number of clusters ($r = 0.994^{**}$), number of pods per cluster ($r = 0.967^{**}$), number of pods per plant ($r = 0.996^{**}$), number of seeds per pod ($r = 0.947^{**}$), number of seeds per plant ($r = 0.999^{**}$), and 100-seed weight ($r = 1.006^{**}$). It was negatively genotypic correlated with days to flowering ($r = -0.912^{**}$) and days to maturity ($r = -0.930^{**}$).

The genotypic correlation coefficients were generally higher in magnitude than their corresponding phenotypic correlations, indicating that the observed associations are primarily genetic rather than environmental. This also suggests that environmental influence had a minimal effect on the expression of these traits.

At the genotypic level, seed yield per plant was highly and positively correlated with number of primary branches per plant ($r_g = 0.930^{**}$), secondary branches ($r_g = 1.000^{**}$), number of leaves ($r_g = 0.910^{**}$), clusters per plant ($r_g = 0.994^{**}$), pods per cluster ($r_g = 0.967^{**}$), pods per plant ($r_g = 0.996^{**}$), seeds per pod ($r_g = 0.947^{**}$), seeds per plant ($r_g = 0.999^{**}$), and 100-seed weight ($r_g = 1.006^{**}$).

These strong associations imply that these yield components are governed by closely linked or pleiotropic genes.

Negative and significant genotypic correlations were observed for days to flowering initiation ($r_g = -0.912^{**}$) and days to maturity ($r_g = -0.930^{**}$) with seed yield per plant, similar to phenotypic correlations. This reinforces the idea that early-flowering and early-maturing mutants possess greater yield potential, likely due to better partitioning of assimilates towards reproductive structures.

The close correspondence between phenotypic and genotypic correlation values for most of the traits suggests that the environmental influence was minimal, indicating that selection based on phenotype for these traits would be effective and reliable.

Comparable observations were made by Kumar et al. (2022) and Kumari et al. (2023) in *Vigna radiata*, who also reported higher genotypic correlations than phenotypic ones for yield-contributing traits such as number of branches, pods, and seed weight.

Evaluation of heritability (narrow sense), genotypic coefficient of variation (GCV), phenotypic coefficient of variation (PCV) and genetic advance (GA) in the M_3 generation of greengram mutants revealed several important insights for breeding and selection.

3.2. Estimation of Heritability and Genetic Advance

The estimates of heritability (narrow sense), genetic advance (GA), genotypic coefficient of variation (GCV), and phenotypic coefficient of variation (PCV) for various quantitative traits in greengram (*Vigna radiata* L.) mutants of the M_3 generation are presented in Table 3.

Table 4. Estimation of heritability, Genotypic and Phenotypic Coefficient of Variations, Genetic Advance and Genetic Advance Value % means for yield and yield related traits in greengram mutant in M_3 generation.

| S. No | Characters | Heritability (%) | Genotypic Coefficient of Variations | Phenotypic Coefficient of Variations | Genetic Advance | Genetic Advance Value % Means |
|-------|------------|------------------|-------------------------------------|--------------------------------------|-----------------|-------------------------------|
| 1 | Pl. Ht(cm) | 99.14 | 34.297 | 34.445 | 31.733 | 70.347 |



| S. No | Characters | Heritability (%) | Genotypic Coefficient of Variations | Phenotypic Coefficient of Variations | Genetic Advance | Genetic Advance Value % Means |
|-------|-----------------|------------------|-------------------------------------|--------------------------------------|-----------------|-------------------------------|
| 2 | No. of P.Br./Pl | 97.864 | 39.19 | 39.615 | 5.513 | 79.864 |
| 3 | No. of S.Br./Pl | 87.011 | 28.637 | 30.7 | 1.563 | 55.028 |
| 4 | Days to F.Ini. | 98.022 | 10.679 | 10.786 | 11.674 | 21.78 |
| 5 | Days to P.M | 85.276 | 5.02 | 5.436 | 8.738 | 9.549 |
| 6 | No. L/Pl | 97.275 | 34.035 | 34.508 | 11.643 | 69.149 |
| 7 | No. Cl/Pl | 93.48 | 30.803 | 31.859 | 2.536 | 61.35 |
| 8 | No. P/Cl | 86.61 | 20.091 | 21.589 | 1.289 | 38.518 |
| 9 | No. P/Pl | 97.306 | 47.662 | 48.317 | 14.013 | 96.852 |
| 10 | No. S/PD | 68.639 | 8.456 | 10.207 | 1.36 | 14.432 |
| 11 | No S/Pl | 94.627 | 53.833 | 55.34 | 152.105 | 107.876 |
| 12 | 100 S. Wt(g) | 64.931 | 3.679 | 4.566 | 0.207 | 6.107 |
| 13 | S.Y(g)/Pl | 94.393 | 57.454 | 59.136 | 5.58 | 114.99 |

Pl. Ht(cm)- Plant Height (cm); No. of P.Br./Pl- Number of Primary Branches per Plant; No. of S.Br./Pl- Number of Secondary Branches per Plant; Days to F.Ini.- Days to Flower Initiation; Days to P.M- Days to Pod Maturity; No. L/Pl- Number of Leaves per plant; No. Cl/Pl- Number of Clusters per plant; No. P/Cl- Number of Pods per Cluster; No. P/Pl- Number of Pods per Plant; No. S/PD- Number of Seeds per Pod; No S/Pl- Number of Seeds per Plant; S.Y(g)/Pl- Seed Yield (g/pl); 100 S. Wt(g)- 100 Seed Weight (g).

High heritability coupled with high genetic advance was observed for plant height, number of primary and secondary branches per plant, number of pods per plant, number of seeds per plant, and seed yield per plant, which indicates that these traits are predominantly controlled by additive gene effects. Therefore, selection based on these traits would be effective for yield improvement in greengram mutants.

In contrast, traits with moderate heritability and low genetic advance such as 100-seed weight and days to maturity suggest the influence of non-additive gene action or environmental factors, requiring further selection cycles or hybridization for improvement.

Table 5. High Heritability and High Genetic Advance Traits in greengram mutants in M3 generation.

| S. No | Characters | Heritability (%) | Genotypic Coefficient of Variations | Phenotypic Coefficient of Variations | Genetic Advance | Genetic Advance Value % Means |
|-------|-----------------|------------------|-------------------------------------|--------------------------------------|-----------------|-------------------------------|
| 1 | Pl. Ht (cm) | 99.14 | 34.297 | 34.445 | 31.733 | 70.347 |
| 2 | No. of P.Br./Pl | 97.864 | 39.19 | 39.615 | 5.513 | 79.864 |
| 3 | Days to F.Ini. | 98.022 | 10.679 | 10.786 | 11.674 | 21.78 |
| 4 | No. L/Pl | 97.275 | 34.035 | 34.508 | 11.643 | 69.149 |
| 5 | No. P/Pl | 97.306 | 47.662 | 48.317 | 14.013 | 96.852 |

Pl. Ht(cm)- Plant Height (cm); No. of P.Br./Pl- Number of Primary Branches per Plant; Days to F. Ini.- Days to Flower Initiation; No. L/Pl- Number of Leaves per plant; No. P/Pl- Number of Pods per Plant.

Traits such as plant height (99.14 %), number of primary branches per plant (97.86 %), number of leaves per plant (97.27 %) and number of pods per plant (97.30 %) exhibited very high heritability associated with high



genetic advance as percent of mean (GAM) (70.347 %, 79.864 %, 69.149 %, 96.852 % respectively). This combination is indicative of predominantly additive gene action, minimal environmental influence and good scope for improvement through selection.

Similar trends have been reported in recent studies on mungbean/greengram, where high heritability along with high expected genetic gain suggested that direct selection would be effective.

Therefore, for these traits in the mutant population, early generation selection can be reliably deployed. The high GCV and PCV for these traits further reflect ample variability in the population. For example, seeds per plant had GCV ~53.83 % and PCV ~55.34 %, with GAM ~107.876 %, pointing to a broad developmental baseline and high selection potential.

3.2.1. Moderate to Low Heritability & Genetic Advance Traits

In contrast, traits such as 100-seed weight (heritability ~64.93 %, GAM ~6.11 %), days to physiological maturity (heritability ~85.28 %, GAM ~9.55 %) and number of seeds per pod (heritability ~68.64 %, GAM ~14.43 %) exhibited moderate to low genetic advance despite moderate to high heritability. This suggests that although the traits are inheritable, additive genetic variance is limited or the trait is under considerable environmental or non-additive genetic control (Khan et al., 2023). In such cases, selection may be less efficient and may require heterosis breeding or recurrent selection to exploit non-additive variance.

3.3. Relationship between GCV and PCV

Across traits, PCV values were marginally higher than GCV values which implies some environmental influence, but the gap was not very wide for most traits. For traits with very high heritability and high genetic advance, the narrow gap between GCV and PCV suggests that environment played a minor role and the observed variation is largely genetic. This pattern is in line with observations in recent legume-mutant studies (Singh & Roy, 2021) where GCV/PCV differences were small for traits with high heritability.

3.4. Implications for Breeding in Greengram Mutants

Because of the high heritability and genetic advance demonstrated by the key yield-contributing traits (pods per plant, seeds per plant, primary branches, leaves), these should be prioritized in selection programs aimed at improving yield in M_3 generation mutants. The strong genetic control implies that phenotype-based selection would likely be effective and can lead to stable improvement. Conversely, traits with low genetic advance (e.g., 100-seed weight) may need to be improved by other methods-for example through hybridization or multiple cycles of selection to accumulate favorable alleles.

Even though many traits show favorable genetic parameters, it is advisable to validate these findings across environments (multi-location trials) because the current estimates are from a single context (M_3 generation mutants). Gene \times environment interactions could still influence expression, especially for traits with moderate heritability. Also, studying the nature of gene action (additive vs dominance) via combining ability or molecular marker-based analysis can further guide breeding strategy (Patel et al., 2024). Some research works have been carried out in black gram using gamma rays, combinations with EMS other chemical alkalinizing agents to increase the yield and yield contributing traits and good results have been reported. Induced mutations are highly used to create genetic variability with desirable agronomical characters in the plants like black gram as



these are self pollinated crops having less chance of genetic variability (Wani, 2021; Raina et al., 2019; Goyal et al., 2019; Laskar et al., 2018)). Similarly in other crops like chickpea (Raina et al 2019), lentil (Laskar et al 2018), mungbean (Singh, 2009), blackgram (Sonu Goyal et al 2020).

IV. CONCLUSION

The present investigation demonstrated that EMS-induced mutagenesis effectively generated substantial genetic variability in greengram, providing valuable scope for yield improvement. Traits such as plant height, number of primary and secondary branches per plant, number of pods per plant, number of seeds per plant, and seed yield per plant exhibited high heritability coupled with high genetic advance, indicating the predominance of additive gene action and the effectiveness of direct selection. Among these, number of pods per plant and seeds per plant emerged as reliable selection indices for enhancing productivity in advanced mutant generations. Conversely, traits like 100-seed weight and days to maturity, which showed moderate heritability with comparatively lower genetic advance, appear to be influenced by non-additive gene effects and environmental factors, suggesting the need for prolonged selection or hybridization strategies.

ACKNOWLEDGEMENT

The present work is part of Ph.D. thesis work. We acknowledge the Acharya N.G. Ranga Agricultural University and Centre for Plant Molecular Biology, Osmania University, Hyderabad for providing both field and lab facilities to conduct experiments. We acknowledge ICAR-CRIDA, Hyderabad for providing seed material of Greengram germplasm.

REFERENCES

- [1] Allard, R.W. (1960). Principles of Plant Breeding. John Willey and Sons Inc., New York.
- [2] Basavaraj Greene, E.A., Codomo, C.A., Taylor, N.E., Henikoff, J.G., Till, B.J., Reynolds, S. H., Enns, L.C., Burtner, C., Johnson, J.E., Odden, A.R., Comai, L., & Henikoff, S. (2019). Molecular basis of EMS-induced mutations in plants. *Mutation Research/Genetic Toxicology and Environmental Mutagenesis*, 826, 15-25. <https://doi.org/10.1016/j.mrgentox.2019.01.005>
- [3] Burton G.W. (1952). Quantitative inheritance in grasses. 6th International Grassland Congress 1: 277-83.
- [4] Chaudhary, S., Meena, R.K., & Singh, M. (2024). Correlation and path coefficient analysis for yield attributes in greengram (*Vigna radiata* L. Wilczek). *Indian Journal of Genetics and Plant Breeding*, 84(2), 284–291. <https://doi.org/10.31742/IJGPB.84.2.10>
- [5] Falconer, D.S., & Mackay, T.F.C. (2023). Introduction to quantitative genetics (6th ed.). Pearson Education.
- [6] Fisher, et al. Statistical tables for biological, agricultural and medical research. (1963). 6th Edition. Hafner Press. New York.
- [7] Gadakh S.R., M.S. Shinde, A.R. Gaikwad and V.R. Patil. (2013). Effect of genotypes and phenological stages on green cane yield, brix and juice yield in sweet sorghum, *Journal of Academia and Industrial Research* Vol. 1(10).
- [8] Gadakh, S.S., Dethle, A.M., Kathale, M.N., & Kahate, N.S. (2013). Genetic diversity for yield and its component traits in green gram [*Vigna radiata* (L.) Wilczek]. *Journal of Crop & Weed*, 9(1), 106–109.
- [9] Goyal, S., Wani, M.R., Laskar, R.A., Raina, A., & Khan, S. (2019). Assessment on cytotoxic and mutagenic potency of gamma rays and EMS in *Vigna mungo* (L.) Hepper. *Biotechnologia Vegetal*, 19, 193-204.
- [10] Goyal, S., Wani, M. R., Laskar, R. A., Raina, A., & Khan, S. (2019). Assessment on cytotoxic and mutagenic potency of gamma rays and EMS in *Vigna mungo* (L.) Hepper. *Biotechnology Vegetal*, 19, 193-204.
- [11] Johnson, H.W., H.F. Robinson and R.E. Comstock, (1955). Estimation of genetic and environmental variability in soybeans. *Agron. J.*, 47: 314-318.
- [12] Kaur, M., Gupta, R., & Singh, G. (2023). Nutritional composition and biofortification potential of mungbean (*Vigna radiata* L. Wilczek). *Frontiers in Plant Science*, 14, 11342. <https://doi.org/10.3389/fpls.2023.011342>
- [13] Khan, M. A., Begum, T., & Ali, S. (2023). Dose optimization of EMS for mutation induction in pulses. *Agricultural Science Digest*, 43(1), 22-29. <https://doi.org/10.18805/ag.D-5402>
- [14] Konda Gajraj Singh, P.K.Sareen and R.P. Saharah. (2000). Induced chlorophyll and morphological mutations in mungbean. *Indian J. Genet.*, 60: 391-393.
- [15] Kumar, M., Patel, D., & Singh, R. (2022). Mutagenic sensitivity and effectiveness of EMS in chickpea. *Indian Journal of Genetics and Plant Breeding*, 82(4), 560–568.
- [16] Kumari, S., Kumar, R., Chouhan, S., & Chaudhary, P.L. (2023). Evaluation of genetic variability and inter-relationship of yield attributes in mungbean mutants. *Frontiers in Genetics*, 14, 1208976.
- [17] Laskar, R.A., Wani, M.R., Raina, A., Amin, R., & Khan, S. (2018). Morphological characterisation of gamma rays induced multi podding mutant (mp) in lentil cultivar Pant L 406. *International Journal of Radiation Biology*, 94, 1049–1053.
- [18] Patel, S., Jha, A., & Kumar, R. (2024). Gene × environment effects on yield traits of *Vigna radiata* mutants: a molecular approach. *Plant Breeding Today*, 39(1), 23-31.
- [19] Raina, A., Khan, S., Wani, M.R., Laskar, R.A., & Mushtaq, W. (2019). Chickpea (*Cicer arietinum* L.) cytogenetics, genetic diversity and breeding, *Legumes* (pp. 53-112).



- [20] Sidramappa, C.R. Konda and Shobharani, M. (2019). Agricultural Research Station, Bidar, UAS, Raichur, India. Int. J. Curr. Microbiol. App. Sci. 8(12): 2991-2994. doi: <https://doi.org/10.20546/ijcmas.2019.812.347>
- [21] Singh, R. & Roy, D. (2021). Assessment of genetic advance and heritability in induced mutants of greengram. Journal of Legume Research, 44(4), 345-352.
- [22] Singh, S.K., Singh I., Singh B.B. and Singh, O. (2009): Correlation and path coefficient studies for yield and its components in Mungbean (*Vigna radiata* (L.) Wilczek). Legum. Res., 32 (3): 180-185.
- [23] Sivasubramanian, S., & Madhava Menon, P. (1973). Genotypic and phenotypic variability in rice. Madras Agriculture Journal. 60(9-12), 1093-1096.
- [24] Sonu Goyal, Mohammad Rafiq Wani, Rafiul Amin Laskar, Aamir Raina, Samiullah Khan. (2020). Mutagenic Effectiveness and Efficiency of Individual and Combination Treatments of Gamma Rays and Ethyl Methanesulfonate in Black Gram (*Vigna mungo* L. Hepper). Advances in Zoology and Botany 8(3): 163-168.
- [25] Wani, M. R. (2021). Comparative biological sensitivity and mutability of chemo-mutagens in lentil (*Lens culinaris* Medik). Legume. Research, 44, 26-30. <https://doi.org/10.18805/LR-4058>.

AUTHOR'S PROFILE

First Author

V. Rohini, Assistant Professor of Biotechnology, Department of Biotechnology, Government Degree College Siddipet, Telangana, India.

Second Author

K. Srinivas Naik, Centre for Plant Molecular Biology, Osmania University, Hyderabad, Telangana, India.

Third Author

Deepika Dasyam, Centre for Plant Molecular Biology, Osmania University, Hyderabad, Telangana, India.

Fourth Author

Sudhakar Poda, Department of Biotechnology, Acharya Nagarjuna University, Nagarjuna Nagar, Guntur, Andhra Pradesh, India